

# Benthic Communities in *Spartina alterniflora*– and *Phragmites australis*– Dominated Salt Marshes in the Hackensack Meadowlands, New Jersey\*

Catherine E. Yuhas,<sup>1</sup> Jean Marie Hartman,<sup>2</sup> and Judith S. Weis<sup>3</sup>

<sup>1</sup> New Jersey Sea Grant College Extension Program, New York-New Jersey Harbor Estuary Program Office, 290 Broadway, 24th Floor, New York, NY 10007 (address at time of research: New Jersey Institute of Technology/Rutgers University, Federated Graduate Program in Environmental Science, Newark, NJ 07102);  
cathy@harborestuary.org

<sup>2</sup> Rutgers University, 93 Lipman Drive, New Brunswick, NJ 08901;  
jhartman@rci.rutgers.edu

<sup>3</sup> Department of Biological Sciences, Rutgers University, 411 Boyden Hall, Newark, NJ 07102; jweis@andromeda.rutgers.edu

## Abstract

*Phragmites australis* is invasive in Atlantic coastal salt marshes and often replaces the native cordgrass, *Spartina alterniflora*. Our research focused on benthic communities found in natural and mitigated *P. australis* and *S. alterniflora* salt marshes at two sites, Sawmill Creek (natural sites) and Mill Creek (mitigation sites) in the Hackensack Meadowlands, New Jersey. The area sampled at the natural sites consisted of adjacent stands of *P. australis* (P) and *S. alterniflora* (S) separated by a tidal creek. At Mill Creek, samples were collected at a 12-year-old mitigation site consisting of *S. alterniflora* (M-12) and a newly mitigated site (M-0) where site regrading had just taken place. Benthic samples were taken at the creek bank and the edge of vegetation for all sites.

Data on salinity levels and textural and structural sediment characteristics were collected at each site. Our data indicate that both the *P. australis* and *S. alterniflora* sites support diverse benthic communities, although there were differences in diversity and composition in the communities found among the different types of grasses. The average abundance of benthic invertebrates at the Sawmill Creek natural sites ranged from about 35,000 per square meter (/m<sup>2</sup>) to over 240,000/m<sup>2</sup>. Taxon richness was ~10 at P and ~7 at S and the *P. australis* site had more common taxa present (1.0% of the total abundance). Average abundance of benthic invertebrates at the Mill Creek mitigation sites ranged from about 130,000/m<sup>2</sup> to 3 million/m<sup>2</sup>. Taxon richness was ~10 at M-12 and ~7 at M-0. This study

---

\* Published online December 8, 2005

suggests that significant change has not occurred at M-12 in the low marsh since the commencement of the mitigation in 1988.

**Keywords:** Benthic invertebrates; estuarine ecology; Hackensack Meadowlands; low marsh; *Phragmites australis*; salt marsh; *Spartina alterniflora*; tidal estuary

## Introduction

*Spartina alterniflora* (salt marsh cordgrass) dominates the low marsh zone of tidal marshes on the East Coast of the U.S. (Teal, 1962; Bertness, 1991). *Spartina alterniflora* provides habitat for salt marsh species, including a food source for benthic invertebrates and insects (Teal 1962; Van Dolah, 1978; Healy & Walters, 1994; Kneib, Ne well, & Hermeno, 1997; Able & Hagan, 2000; Graca, Newell, & Kneib, 2000). Another salt marsh grass, *Phragmites australis* (common reed) is an invasive species usually found on high marshes, but it has been moving into low marshes and replacing *S. alterniflora* (Fell et al. 1998; Angradi, Hagan & Able, 2001; Windham & Lathrop, 1999; Weinstein & Balletto, 1999). *Phragmites australis* changes the marsh physically, hydrologically, and chemically (Angradi et al., 2001; Windham & Lathrop, 1999), and this can affect the utilization of the marsh by fish, birds, and other animals. *Phragmites australis* has been replacing native vegetation on the Atlantic and Gulf coasts since the early 1900s (Weinstein & Balletto, 1999; Fell et al., 1998; Angradi et al., 2001; Windham & Lathrop, 1999). Marsh managers have responded by trying to decrease the dominance of *P. australis* on salt marshes. Numerous restoration projects have been undertaken in which *P. australis* was removed and *S. alterniflora* replanted. However, there have been few direct comparisons of the

relative level of function of marshes before and after restoration.

Recent studies on *P. australis* have found it to be an ecologically functional habitat for salt marsh inhabitants such as nekton (fishes and swimming decapod crustaceans) and benthic invertebrates (Rilling, Fell, & Warren, 1998; Meyer, Johnson, & Gill, 2001; Angradi et al., 2001; Fell et al., 1998). There have also been recent studies, using stable isotopes, which have found *P. australis* to be a food source for fish (Wainright, Weinstein, Able & Currin, 2000; Weinstein et al., 2000). However, other studies have found *P. australis* to be poor nursery habitat for the mummichog, *Fundulus heteroclitus* (Able and Hagan 2000; Raichel, Able, & Hartman, 2003).

Benthic invertebrates are vital to a functioning salt marsh ecosystem. They are a food source for many salt marsh inhabitants (Kneib, 1988; Fell et al., 1988; Sarda, Foreman & Valiela, 1995) and important components of the estuarine food web (Ishikawa, 1989). Benthic communities may be affected by the invasion of *P. australis*, though studies to determine the nature of the effects have been contradictory. Fell et al. (1998) conducted research along the Connecticut River and found that *P. australis* salt marshes were functionally equivalent to non-*P. australis* salt marshes. Four high-marsh macroinvertebrates (*Orchestia grillus*, *Philoscia vittata*, *Melampus bidentatus*, and *Succinea* species)—all of them prey species for *F. heteroclitus*—were the focus of this study, and they were found in both marsh types. However, Angradi et al. (2001) found that a *Spartina* marsh had greater production of benthic infauna than a *Phragmites* marsh. Overall abundance of benthic invertebrates was higher, and taxon richness was significantly higher in the *Spartina* marsh than the *Phragmites*

marsh at all sampling positions and dates. Dominance by the three most abundant taxa (Oligochaeta, Nematoda, and *Manayunkia aestuarina*) was greater in the *Phragmites* marsh (> 85%) than the *Spartina* marsh at most of the sampling positions, indicating a lower benthic diversity in the *Phragmites* marsh (Angradi et al., 2001). Posey, Alphin, Meyer, and Johnson (2003) found only minor differences between *Spartina* and *Phragmites* marshes in the Chesapeake Bay, although most species were slightly more abundant in *Spartina*.

Benthic invertebrates can be used to assess whether or not salt marsh restorations are functioning normally as ecosystems. In order to create a functioning ecosystem, a salt marsh restoration should include the reintroduction of vegetation and the duplication of the nekton and benthos, along with other environmental factors of the marsh (Packard & Stiverson, 1976; Allen et al., 1994; Sacco Seneca & Wentworth, 1994). Studies comparing natural and restored marshes have found similarities and differences with regard to fauna (Minello & Webb, 1997; Minton, 1999; Craft, Broome & Sacco, 1998; Havens, Varnell & Bradshaw, 1995; LaSalle, Landin & Sims, 1991; Sacco et al., 1994; Moy & Levin 1991). However, it's uncertain how many years it takes for benthic communities in restored or created marshes to become comparable to those in natural marshes. Created marshes are inhabited by opportunistic benthic species, which develop according to the sediment, hydrodynamics, and vegetation of the marsh (Posey, Alphin & Powell, 1997). The more recently created marshes studied by Posey et al. (1997) had more polychaetes than the older created marshes, which had more oligochaetes and amphipods. Packard and Stiverson (1976) suggest that a *Spartina* marsh restored on dredge

spoils will eventually sustain a detritus-based community, which is a major food source for benthic invertebrates.

In a few studies, restored salt marshes have been deemed to have characteristics similar to natural marshes (Posey et al., 1997). Some studies indicate that the benthic communities in natural and restored marshes are similar. For example, LaSalle et al (1991) found that benthic communities, along with fish and shellfish, of a natural marsh that developed on dredged material were similar in species composition and abundance to those of natural marshes.

Our study has two objectives. The first is a comparison of the benthic communities of a natural *Spartina* marsh and a *Phragmites* marsh. The second is a comparison of two restored marshes of different ages (> 10 years, < 1 year).

## Methods

### Study Sites

The Hackensack Meadowlands District covers 32 square miles in Bergen and Hudson counties of New Jersey (Figure 1). It is uncertain when *P. australis* first appeared here, but published sightings of the plant in New Jersey date back to the 1800s (Willis, 1877; Britton, 1889; Harshberger & Burns, 1919). In our study, we conducted a comparison of natural marshes dominated by *S. alterniflora* and *P. australis* at Sawmill Creek (natural sites, Figure 1) and an assessment of mitigated intertidal marshes at Mill Creek (mitigation sites, Figure 1).

### Sawmill Creek Natural Sites

The study sites located at Sawmill Creek run along the southern section of the Hackensack River in Lyndhurst, New Jersey. This area is a natural salt marsh that was formed in 1950 after a tide-gate

breach (Kraus & Kraus, 1988). The known salinity range at this site is 6.9 to 15.7 parts per thousand (ppt)—mesohaline—and the tidal range is approximately 1.5 meters. *Phragmites australis* is the dominant vegetation at the natural sites, but extensive marshes of *S. alterniflora* can be found as well. We sampled from microhabitats located in adjacent stands of *P. australis* (P) and *S. alterniflora* (S) separated by a tidal creek (Figure 2).

### Mill Creek Mitigation Sites

The mitigated marshes were located at Mill Creek, in Secaucus, New Jersey (Figure 1), an area dominated by *P. australis*. One of the marshes was mitigated in 1988, 12 years before this study, while the other was undergoing mitigation at the time of the sampling. The 12-year-old site (M-12, Figure 3), located along the northern part of the Hackensack River (Hackensack Meadowlands Development Commission, HMDC, 2000), covers 63 acres and was mitigated by Hartz Mountain Company. Mitigation consisted of removing *P. australis*, decreasing site elevation to enhance daily tidal inundation, planting *S. alterniflora*, and site monitoring (TAMS, 1990). This site is completely surrounded by industry, highways, and numerous forms of urbanization (TAMS, 1990; HMDC, 2000). The tidal range at M-12 is approximately 1.5 meters, and the salinity range is 0.5 to 7.0 ppt—oligohaline (Kraus & Kraus, 1988).

The newly mitigated site (M-0, Figure 4), located behind the Mill Creek Mall, in Secaucus, covers an area of 140 acres and was overseen by the New Jersey Meadowlands Commission (formerly the Hackensack Meadowlands Commission). Mitigation began here in 1998 and was conducted to eliminate *P. australis*, increase tidal inundation, and create a low marsh system (HMDC, 2000). At the time of

sampling at the M-0 site, the mitigation was still under way, and all that remained was bare substrate, some *P. australis*, and *Pluchea purpurascens*, an annual salt marsh fleabane that flowers between late summer and fall (Newcomb, 1977).

### Organic Matter and Particle Size

We took sediment samples to a depth of 5 centimeters (cm) in September 1999 using a PVC corer. Three replicates were made at each of the four sampling stations. Samples were kept in a cold room prior to processing. The samples were initially sieved wet through a 4.75-millimeter (mm) sieve to collect any pebbles, stones, and large pieces of organic material. We determined the following sediment characteristics: percentage organic matter and percentage silt, sand, and clay.

To determine organic content, five grams of wet sediment were placed in a ceramic crucible, weighed, and placed in a 105°C oven for 16 hours. Samples were then reweighed to determine the dry weight, placed in a hood, and put in a 440°C muffle furnace for 16 hours. Samples were then cooled in the hood and reweighed in order to determine ash dry weight. The percentage of organic matter was calculated using the following equation:  $(\text{Dry Weight} - \text{Ash Weight}) / \text{Dry Weight} \times 100$ .

To determine particle size, the remaining sediment samples were air dried in a university greenhouse. Pieces greater than 3.175 mm in diameter were removed from the sediment after it was crushed with a mortar and pestle. The sediment was analyzed using the LaMotte Soil Texture Unit (code 1067, LaMotte Co., Chestertown, MD), which provides a volumetric calibration of sand, silt, and clay through sedimentation in an aqueous solution.

### **Benthic Sampling**

We collected benthic samples in the natural (S and P) and mitigation (M-0 and M-12) sites at two low-marsh microhabitats: (1) the creek bank and (2) the edge of each type of vegetation on the marsh surface. At the natural sites, the creek bank sampled was a steep area coming off the vegetated marsh surface into an emergent area in the intertidal zone. At the mitigation sites, the creek bank sampled was a gently sloping emergent area in the intertidal zone coming off the vegetated (M-12) or nonvegetated (M-0) marsh surface. At M-0, the “edge of vegetation” was estimated by examining vegetation surrounding the site.

Benthic samples were taken at all sites with a 3.9-cm-diameter PVC core sampler. The cores were taken to a depth of 5 cm at each sampling station. Cores were taken at this relatively shallow depth because it has been shown that in stressed marshes (for example, those that are contaminated or have low dissolved oxygen), the large, deep-burrowing invertebrates disappear, and the community becomes dominated by small animals that live close to the sediment surface (Warwick, 1993). Weis, Skurnick, and Weis (2004) have found an absence of larger, deeper-dwelling fauna in the Meadowlands.

We collected three replicates at all sampling sites. Creek-bank samples were collected once a month at low tide from June to September 1999. Edge-of-vegetation samples were collected from July to September 1999. Surface-water salinity at each site was measured with a refractometer.

### **Biota**

Unsieved benthic samples (mud and organisms) were preserved in 20% formalin in the field. After one to

two weeks, samples were transferred to 70% ethanol with rose bengal dye for staining of the benthic invertebrates. The samples were sieved in a 0.3-mm sieve, sorted, and identified in a petri dish to the lowest possible taxonomic level to determine taxa richness, overall abundance, and species composition (Weiss, 1995). The numbers of nematodes were estimated for a site when an average of  $\geq 100$  nematodes were found in each sample. The petri dishes utilized were counted and multiplied by the mean number of nematodes in the first two petri dishes sorted for that particular site. For this study, composition comparisons focused on those taxa that were classified common ( $\geq 1.0\%$  of the total abundance) and uncommon ( $> 0.1\%$  but  $< 1.0\%$ ) (LaSalle & Rozas, 1991).

### **Statistical Analysis**

We analyzed all the benthic samples using the Statistical Analysis System (SAS) software (Version 8). Multiple analysis of variance (MANOVA) and the Student-Newman-Keuls (SNK) tests were used to analyze the monthly benthic samples ( $p < 0.05$ ). The percentage of organic matter and the percentage of silt, sand, and clay were analyzed by running analysis of variance (ANOVA). All data were tested for normality, and any abundance data with abnormal distribution were log-base-10 transformed. A commonly used measure of diversity, the Shannon-Wiener Index, was calculated using Multivariate Statistical Package (MVSP) software (Version 3.12c).

## Results

### Sawmill Creek Natural Sites

#### Organic Matter, Particle-Size Distribution, and Salinity

We found a significant difference between the percentage of organic matter in the creek-bank sediment samples from S (3.08%) and P (1.58%) (Table 1 and Table 2,  $p = 0.0003/df = 2$ ). There was no significant difference in percentage of organic matter at the edge of the vegetation among the natural sites (Table 1 and Table 2, S: 2.18%; P: 2.28%). However, the mean percentage of organic matter was significantly different between the creek bank and the edge of the vegetation for both *P. australis* ( $p = 0.0656/df = 1$ ) and *S. alterniflora* ( $p = 0.0251/df = 1$ ).

We found no significant differences in percentage of silt in the creek-bank samples at the natural sites. The sediment was significantly coarser at S than P on the creek bank since S had more sand present (Table 1 and Table 2,  $P = 0.0038/df = 2$ ). There were fewer fine particles (less clay content) at S than P (Figure 5, Table 2,  $p = 0.0128/df = 1$ ). We found no significant difference between sites in particle-size distribution at the edge of the vegetation.

At both natural sites, the salinity was 8 ppt in June, 15 ppt in July, 20 ppt in August, and 4 ppt in September. The increase in salinity from June to August was due to a drought during the summer of 1999. The drastic salinity decrease in September was due to rainfall associated with Hurricane Floyd on September 16, 1999.

#### Abundance, Richness, and Composition

We found no significant differences in the total number of organisms over all months sampled on the creek bank for the natural sites (Figure 6, Table 3 and

Table 4,  $p > 0.05/df = 1$ ). Mean abundance at the edge of the vegetation was not significantly different for July and August (Figure 6, Table 4), but during September, P had significantly less abundance than S ( $p = 0.0212/df = 1$ ). The natural sites showed no significant difference for July, August, and September between creek-bank and vegetation-edge habitat types for S or for P.

There was no significant difference in taxa between S and P at the creek bank for all months sampled. The mean taxa richness at the edge of the vegetation did show significant differences for some months (Figure 7, Table 3 and Table 4,  $p < 0.05/df = 1$ ). During August, P had a significantly higher number of taxa than S at the edge of the vegetation (Table 4,  $p = 0.0255/df = 1$ ). However, there was no significant difference between S and P at the creek bank for July and September. In August, the creek bank of S had significantly higher mean taxa richness ( $p = 0.0241/df = 1$ ) compared to P.

Presence (+) and absence (–) of the 25 taxa found over the four months at the sites is documented in Table 5. Three types of meiofauna (Nematoda, Copepoda, and Ostracoda) and several macrofaunal taxa dominated the collections. We identified ten common taxa and eight uncommon taxa at P (Table 6). Eight common taxa were found at S, along with six uncommon taxa (Table 6). Oligochaeta (24.4%) had the highest percent composition at P, while *Manayunkia aestuarina* (33.0%) had the highest percent composition at S.

#### Shannon-Wiener Diversity Index

We found a significant difference between taxa diversity on the creek banks of the different sites for June. In this month, P was significantly more diverse than S ( $p = 0.0347/df = 1$ ), but during July, August,

and September, there was no significant difference ( $p > 0.05/df = 1$ ) between P and S (Table 3 and Table 4). At the edge of the vegetation, P had significantly more diversity than S in August ( $p = 0.0371/df = 1$ ), but not in July and September ( $p > 0.05/df = 1$ ).

The taxa diversity between the two habitats at S was significantly different in July ( $p = 0.0218/df = 1$ ) and August ( $p = 0.0189/df = 1$ ), when the creek bank was more diverse than the edge of the vegetation (Table 4). For September, there was no significant difference between the two habitats at S. At P, there was no significant difference in July and September ( $p > 0.05/df = 1$ ), but in August the creek bank was significantly more diverse than the edge of the vegetation ( $p = 0.0081/df = 1$ ).

#### Mill Creek Mitigation Sites

##### Organic Matter, Particle Size Distribution and Salinity

Organic matter at the creek bank for M-0 was calculated to be 1.69% compared to 1.33% at M-12 (Table 1 and Table 2,  $p = 0.0027/df = 2$ ). There was no significant difference at the edge of the vegetation among the sites (M-12: 2.02%; M-0: 2.31%). For both sites, the mean percentage of organic matter was significantly higher at the edge of the vegetation habitat than the creek bank (M-12:  $p = 0.0088/df = 1$ ; M-0:  $p = 0.0075/df = 1$ ).

We found no differences in silt, sand, and clay percentages (Table 1 and Table 2; Figure 8) between the creek banks of both mitigation sites. In addition, no difference was discernable at the edge of the vegetation between the sites.

Salinity at the mitigation sites (M-12) was 3 ppt in June, 10 ppt in July, 10 ppt in August, and 2 ppt in September. The mitigation sites at Mill Creek,

following a pattern similar to the natural sites at Sawmill Creek, decreased in salinity in September in concurrence with the rainfall of Hurricane Floyd.

##### Abundance, Richness, and Composition

We found no significant difference in the total number of organisms on the creek bank of the mitigation sites for all months sampled (Table 3 and Table 4, Figure 9). The mean abundance at the edge of the vegetation was not significantly different between the sites for July (Figure 9, Table 4,  $p = 0.8001/df = 1$ ). However, during August ( $p = 0.0133/df = 1$ ) and September ( $p < 0.0001/df = 1$ ), abundance at M-12 was significantly greater than at M-0. Abundance was greater at the marsh edge than the creek-bank habitats for July at M-12 ( $p = 0.035/df = 1$ ); and it was greater at the creek bank than the marsh edge for August ( $p = 0.0024/df = 1$ ) and September ( $P < 0.0001/df = 1$ ) at M-0 (Figure 9, Table 4).

In August and September, the taxa richness on the creek bank at M-0 was lower than at M-12 (Table 3 and Table 4, Figure 10, August:  $p = 0.0004/df = 1$ ; September:  $p = 0.0058/df = 1$ ). During July, M-12 had more taxa present than M-0 at the edge of the vegetation (Table 3 and Table 4, Figure 10,  $p = 0.0048/df = 1$ ). The mean taxa richness for M-12 was significantly higher at the creek bank in August ( $p = 0.0158/df = 1$ ) and September ( $p = 0.0335/df = 1$ ) than at the edge of the vegetation.

Refer to Table 5 for presence (+) and absence (–) of the 25 taxa found over four months at all of the sites. At M-12, four common taxa were found along with two uncommon taxa (Table 6). Four common taxa and three uncommon taxa were found at M-0 (Table 6). Nematoda were the most abundant taxa at both M-12 and M-0. The mitigation sites were

heavily dominated by nematodes, which comprised approximately 77%–80% of all taxa.

### Shannon-Wiener Diversity Index

In August, we found that M-12 was more diverse than M-0 at the creek bank (Table 3 and Table 4,  $p = 0.0464/df = 1$ ). During June, July, and September, there was no significant difference between M-12 and M-0. At the edge of the vegetation, M-12 had less diversity than M-0 in August ( $p = 0.0225/df = 1$ ) and September ( $p = 0.0014/df = 1$ ), but during July, M-12 was significantly more diverse than M-0 ( $p = 0.0333/df = 1$ ).

At M-12, benthic taxa diversity was not significantly different between the creek bank and the edge of the vegetation for July, August, or September. The taxa diversity between the two habitats at M-0 was significantly different in August ( $p = 0.0012/df = 1$ ) and September ( $p = 0.0007/df = 1$ ), when the edge of the vegetation was more diverse than the creek bank. For July, there was no significant difference between the two habitats at M-0.

## Discussion

### Sawmill Creek Natural Sites

Angradi et al. (2001) compared the benthic communities in *P. australis* and *Spartina* marshes and found that the *Spartina* marsh had a greater abundance and taxa richness than *P. australis* marsh. However, we found no clear pattern of difference in taxa abundance and richness at *P. australis* and *S. alterniflora* marshes during our study period. As in Angradi et al.'s study, oligochaetes, nematodes, and *Manayunkia aestuarina* were the dominant taxa in our study. And both studies included samples from the creek bank and the edge of the vegetated marsh. However, Angradi et al. sampled from within the

vegetated zone of the marsh, whereas we did not. Since our samples were collected from the creek bank and the marsh edge only, the data may not be reflective of the marsh surface.

There have been studies that have looked at marsh surface and made comparisons between *P. australis* and *S. alterniflora* marshes, including Posey et al. (2003) and Fell et al. (1998). Posey et al. found that the macrobenthic communities in paired *P. australis* and *S. alterniflora* marshes of Chesapeake Bay—an oligohaline to mesohaline environment—were affected by the vegetation type. While they noted few significant differences in the abundance of most individual taxa, they did find a significant overall community trend toward a higher rank abundance of the invertebrates in the *S. alterniflora* marsh compared to the *P. australis* marsh. Fell et al. looked at epibenthic communities in non-*P. australis* and *P. australis* marshes and found them to be equivalent.

### Mill Creek Mitigation Sites

TAMS (1990) evaluated the benthic community in the open-water channels at M-12 and found it to have low diversity. Ten years after this study, our investigations showed that the low diversity still persists. TAMS also found that the benthic community at M-12 consisted of pollution-tolerant organisms, with oligochaetes and hydrobiid gastropods comprising more than 80% percent of the community. In our study, nematodes made up about 80% of the benthic community, and there were only four common taxa found at this site. We conclude that this site is still dominated by few taxa with large abundances.

Our results are similar to those of Kraus and Kraus (1988), who studied mitigation and natural



sites in Sawmill Creek and sampled the mitigated (*S. alterniflora*) and nonmitigated (*P. australis*) sections of Mill Creek. Like us, they found that Sawmill Creek had a greater abundance, higher taxa richness, and higher diversity of benthic invertebrates than the Mill Creek sites, which were dominated by gastropods and nematodes. However, there are two serious obstacles to comparing the studies. First, Kraus and Kraus did not specify the precise locations sampled at Sawmill Creek. Nor did they specify exactly where the Mill Creek control sites were located or which type of vegetation was present there. Second, previous studies have shown that low salinity decreases abundance, taxa richness, and diversity of benthic communities (Levin & Talley, 2000; Boesch, 1972; West & Ambrose, 1992). Insects and oligochaetes usually dominate benthic communities in a low-salinity system, while a high-salinity system is known to favor polychaetes (Levin & Talley). There is evidence for this in our study, which shows that the high-salinity natural sites had considerably more polychaetes than the low-salinity mitigation sites, and that both site types had an abundance of oligochaetes. Oligochaetes were one of the few common taxa found at Mill Creek, while at Sawmill Creek they were one of the many common taxa.

Studies have shown that contaminants and pollutants affect benthic communities (Gray, Clarke, Warwick & Hobbs, 1990; Pocklington & Wells, 1992; Gaston & Young, 1992; Whaley, Garcia & Sy, 1989; Maltby, 1999; Flynn, Wakabara & Tararam, 1998). According to Levin and Talley (2000), a marsh exposed to sewage has a greater abundance of the oligochaete *Monopylephorus rubroniveus* and the amphipod *Talorchestia longicornis*. The mitigation sites at Mill Creek had abundant oligochaetes. A

contamination source that may be affecting the benthic community at the mitigation sites is the sewage-treatment plant along Mill Creek. Kraus and Kraus (1988) reported that the water quality was better at the natural sites than at the mitigation sites. However, during the dates of our study, water quality was better at the mitigation sites than at the natural sites (Center for Information Management, Integration and Connectivity, CIMIC, 1999). But water quality on a single date is far less important than long-term sediment concentration, which would impact benthos. Contamination persists in the mitigation sites' sediments, but an experiment replacing contaminated sediments with uncontaminated ones showed no shift in community composition (Yuhas, 2001).

Overall, this study suggests that significant change has not occurred at the M-12 site at the low marsh since the commencement of the mitigation in 1988. Development of a diverse benthic community may not have occurred because of the low salinity that is representative of an oligohaline area. Changing the vegetation from *P. australis* to *S. alterniflora* may not, alone, be able to alter the environmental conditions (Packard & Stiverson, 1976; Allen et al., 1994; Sacco et al., 1994) at Mill Creek that affect the benthic community.

This study was a small pilot effort. It is consistent in several ways with some previous studies of the Hackensack marsh system. However, it also shows that a more in-depth examination of the complex interactions between salinity, vegetation, time, and contamination is needed to understand the structure and dynamics of the benthic community in the system.

## Acknowledgments

We would like to acknowledge Ed Konsevick and Brett Bragin for facilitating fieldwork, and Lisa Windham for all her help in the field. We would also like to express our gratitude to all the members of the Hartman Lab Group for their assistance and advice. Inga Parker's assistance with editing transformed the manuscript and is much appreciated. We would like to thank the editors and the reviewers for their comments that improved the manuscript. This research was funded by a grant from the New Jersey Meadowlands Commission under the direction of Dr. Jean Marie Hartman and the New Jersey Agriculture Experiment Station.

## Literature Cited

- Able, K.W. & Hagan, S.M. (2000). Effects of common reed (*Phragmites australis*) invasion on marsh surface macrofauna: response of fishes and decapod crustaceans. *Estuaries*, 23, 633–646.
- Allen, E.A., Fell, P.E., Peck, M.A., Gieg, J.A., Guthke, C.R. & Newkirk, M.D. (1994). Gut contents of common mummichogs, *Fundulus heteroclitus* L., in a restored impounded marsh and in natural reference marshes. *Estuaries*, 17, 462–471.
- Angradi, T.R., Hagan, S.M. & Able, K.W. (2001). Vegetation type and the intertidal macroinvertebrate fauna of a brackish marsh: *Phragmites* vs. *Spartina*. *Wetlands*, 21, 75–92.
- Bertness, M.D. (1991). Zonation of *Spartina patens* and *Spartina alterniflora* in a New England salt marsh. *Ecology*, 72, 138–148.
- Boesch, D.F. (1972). Species diversity of marine macrobenthos in the Virginia area. *Chesapeake Science*, 13, 206–211.
- Britton, N.L. (1889). Catalogue of plants found in New Jersey. In Geological Survey of New Jersey, Final report of the state geologist: Vol. II (p. 293). Trenton: John L. Murphy Publishing Company.
- Center for Information Management, Integration and Connectivity (CIMIC). (1999). Meadowlands Environmental Research Institute (MERI). Digital Meadowlands. Hackensack Meadowlands Development Commission Monitoring. Water Quality Program.  
http://cimic.rutgers.edu/hmdc\_public/monitoring/.
- Craft, C.B., Broome, S.W. & Sacco, J.A. (1998, April). Twenty-five years of ecosystem succession on newly created salt-marshes [Abstract]. Paper presented at international conference, Concepts and Controversies in Tidal Marsh Ecology, Vineland, NJ.
- Fell, P.E., Weissbach, S.P., Jones, D.A., Fallon, M.A., Zeppieri, J.A., Faison, E.K., Lennon, K.A., Newberry, K.J. & Reddington, L.K. (1998). Does invasion of oligohaline tidal marshes by reed grass, *Phragmites australis* (Cav.) Trin. ex Steud., affect the availability of prey resources for the mummichogs, *Fundulus heteroclitus* L.? *Journal of Experimental Marine Biology and Ecology*, 222, 59–77.
- Flynn, M.N., Wakabara, Y. & Tararam, A.S. (1998). Macrobenthic associations of the lower and upper marshes of a tidal flat colonized by *Spartina alterniflora* in Cananeia Lagoon Estuarine region (southeastern Brazil). *Bulletin of Marine Science*, 63, 427–442.
- Gaston, G.R. & Young, J.C. (1992). Effects of contaminants of macrobenthic communities in the upper Calcasieu Estuary, Louisiana. *Bulletin of Environmental Contamination and Toxicology*, 49, 922–928.
- Graca, M.A., Newell, S.Y. & Kneib, R.T. (2000). Grazing rates of organic matter and living fungal biomass of decaying *Spartina alterniflora* by three species of salt-marsh invertebrates. *Marine Biology*, 136, 281–289.
- Gray, J.S., Clarke, K.R., Warwick, R.M. & Hobbs, G. (1990). Detection of initial effects of pollution on marine benthos: an example from the Ekofisk and Eldfisk oilfields, North Sea. *Marine Ecology Progress Series*, 66, 285–299.
- Hackensack Meadowlands Development Commission (HMDC). (2000). *Internal report for the National Academy of Sciences Meadowlands Mitigation Project Summary*. Lyndhurst, NJ: author.

- Harshberger, J.W. & Burns, V.G. (1919). The vegetation of the Hackensack Marsh: a typical American fen. *Transactions of the Wagner Free Institute of Science of Philadelphia*, 9, 1–35.
- Havens, K.J., Varnell, L.M. & Bradshaw, J.G. (1995). An assessment of ecological conditions in a constructed tidal marsh and two natural reference tidal marshes in coastal Virginia. *Ecological Engineering*, 4, 117–141.
- Healy, B. & Walters, K. (1994). Oligochaeta in *Spartina* stems: the microdistribution of Enchytraeidae and Tubificidae in a salt marsh, Sapelo Island, USA. *Hydrobiologia*, 278, 111–123.
- Ishikawa, K. (1989). Relationship between bottom characteristics and benthic organisms in the shallow water of Oppa Bay, Miyagi. *Marine Biology*, 102, 265–273.
- Kneib, R.T. (1988). Testing for indirect effects of predation in an intertidal soft-bottom community. *Ecology*, 69, 1795–1805.
- Kneib, R.T., Newell, S.Y. & Hermeno, E.T. (1997). Survival, growth, and reproduction of the salt-marsh amphipod *Uchlorchestia spartinophlia* reared on natural diets of senescent and dead *Spartina alterniflora* leaves. *Marine Biology*, 128, 423–431.
- Kovach, W. (2000). Multivariate Statistical Package (MVSP) (Version 3.12c) [Computer software]. Anglesey, Wales: Kovach Computing Services.
- Kraus, D.B. & Kraus, M.L. (1988). The establishment of a fiddler crab (*Uca minax*) colony on a manmade *Spartina* mitigation marsh, and its effect on invertebrate colonization (Proceedings of the National Wetland Symposium: Mitigation of Impacts and Losses. New Orleans, Louisiana. October 8–10, 1986). *Association of State Wetland Managers (ASWM) Technical Report 3*, 343–348.
- LaSalle, M.W., Landin, M.C. & Sims, J.G. (1991). Evaluation of the flora and fauna of a *Spartina alterniflora* marsh established on dredged material in Winyah Bay, South Carolina. *Wetlands*, 11, 191–208.
- LaSalle, M.W. & Rozas, L.P. (1991). Comparing benthic macrofaunal assemblages of creek bank beds of the spikerush *Eleocharis parvula* (R&S) link and adjacent unvegetated areas in a Mississippi brackish marsh. *Wetlands*, 11, 229–244.
- Levin, L.A. & Talley, T.S. (2000). Influences of vegetation and abiotic environmental factors on salt marsh invertebrates. In M.P. Weinstein & D.A. Kreeger (Eds.), *Concepts and controversies in tidal marsh ecology* (pp. 661–707). Dordrecht, Netherlands: Kluwer Academic Publishers.
- Maltby, L. (1999). Studying stress: the importance of organism-level responses. *Ecological Applications*, 9, 431–440.
- Meyer, D.L., Johnson, J.M. & Gill, J.W. (2001). A comparison of nekton use of *Phragmites australis* and *Spartina alterniflora* marshes in the Chesapeake Bay, USA. *Marine Ecology Progress Series*, 209, 71–83.
- Moy, L.D. & Levin, L.A. (1991). Are *Spartina* marshes a replaceable resource? A functional approach to evaluation of marsh creation efforts. *Estuaries*, 14, 1–16.
- Minello, T.J. & Webb, J.W. Jr. (1997). Use of natural and created *Spartina alterniflora* salt marshes by fishery species and other aquatic fauna in Galveston Bay, Texas, USA. *Marine Ecology Progress Series*, 151, 165–179.
- Minton, M.D. (1999). Coastal wetland restoration and its potential impact on fishery resources in the northeastern United States. *American Fisheries Society Symposium*, 22, 405–420.
- Newcomb, L. (1977). *Wildflower guide*. New York: Little, Brown and Company.
- Packard, G.C. & Stiverson, R.K. (1976). Abundance and production of macroinvertebrates from natural and artificially established salt marshes in North Carolina. *The American Midland Naturalist*, 96, 487–493.
- Pocklington, P. & Wells, P.G. (1992). Polychaetes key taxa for marine environmental quality monitoring. *Marine Pollution Bulletin*, 24, 593–598.
- Posey, M.H., Alphin, T.D. & Powell, and C.M. (1997). Plant and infaunal communities associated with a created marsh. *Estuaries*, 20, 42–47.

- Posey, M.H., Alphin, T.D., Meyer, D.L. & Johnson, J.M. (2003). Benthic communities of common reed *Phragmites australis* and marsh cordgrass *Spartina alterniflora* marshes in Chesapeake Bay. *Marine Ecology Progress Series*, 261, 51–61.
- Raichel, D.L., Able, K.W. & Hartman, J.M. (2003). The influence of *Phragmites* (common reed) on the distribution, abundance, and potential prey of a resident marsh fish in the Hackensack Meadowlands, New Jersey. *Estuaries*, 26, 511–521.
- Rilling, G.C., Fell, P.E. & Warren, R.S. March 26, 1998. *Fish use of brackish high marsh areas in the lower Connecticut River: a comparison of a Phragmites australis–dominated versus a restored marsh* (Final report). Hartford, CT: Connecticut Department of Environmental Protection, Office of Long Island Sound Programs.
- Sacco, J.N., Seneca, E.D. & Wentworth, T.R. (1994). Infaunal community development of artificially established salt marshes in North Carolina. *Estuaries*, 17, 489–500.
- Sarda, R., Foreman, K., & Valiela, I. (1995). Macroinfauna of a southern New England salt marsh: seasonal dynamics and production. *Marine Biology*, 121, 431–445.
- TAMS. (1990). *Comprehensive baseline studies: IR-2 site and off-site mitigation areas: evaluation of Harmon Meadow Western Brackish Marsh Mitigation Area* (Report prepared for Hartz Mountain Industries, Inc.) New York: author.
- Teal, J.M. (1962). Energy flow in the salt marsh ecosystem of Georgia. *Ecology*, 43, 614–624.
- Van Dolah, R.F. (1978). Factors regulating the distribution and population dynamics of the amphipod *Gammarus palustris* in an intertidal salt marsh community. *Ecological Monographs*, 48, 191–217.
- Wainright, S.C., Weinstein, M.P., Able, K.W. & Currin, C.A. (2000). Relative importance of benthic microalgae, phytoplankton and the detritus of smooth cordgrass *Spartina alterniflora* and the common reed *Phragmites australis* to brackish-marsh food webs. *Marine Ecology Progress Series*, 200, 77–91.
- Warwick, R.M. (1993). Environmental impact studies on marine communities: Pragmatical considerationns. *Australian Journal of Ecology*, 18, 63–80.
- Weinstein, M.P. & Balletto, J.H. (1999). Does the common reed, *Phragmites australis*, affect essential fish habitat? *Estuaries*, 22, 793–802.
- Weinstein, M.P., Litvin, S.Y., Bosley, K.L., Fuller, C.M. & Wainright, S.C. (2000). The role of tidal salt marsh as an energy source for marine transient and resident finfishes: A stable isotope approach. *Transactions of the American Fisheries Society*, 129, 797–810.
- Weis, J.S., Skurnick, J. & Weis, P. (2004). Studies of a contaminated brackish marsh in the Hackensack Meadowlands of northeastern New Jersey: benthic communities and metal contamination. *Marine Pollution Bulletin*, 49, 1025–1035.
- Weiss, H.M. (1995). *Marine animals of southern New England and New York* (Bulletin 115 of the State Geological and Natural History Survey of Connecticut). Hartford, Connecticut: Connecticut Department of Environmental Protection.
- West, T.L. & Ambrose, W.G. Jr. (1992). Abiotic and biotic effects on population dynamics of oligohaline benthic invertebrates. In G. Colombo, I. Ferrai, V. Ceccherelli, & R. Rossi (Eds.), *Marine eutrophication and population dynamics* (pp. 189–194). Fredensburg, Denmark: Olsen and Olsen.
- Willis, O.R. (1877). *Catalogue of plants growing without cultivation in the state of New Jersey, with a specific description of all the species of a violet found therein*. New York: A.S. Barnes and Company.
- Whaley, M., Garcia, R., & Sy, J. (1989). Acute bioassays with benthic macroinvertebrates conducted in situ. *Bulletin of Environmental Contamination and Toxicology*, 43, 570–575.
- Windham, L. & Lathrop, R.G. Jr. (1999). Effects of the (common reed) invasion on aboveground biomass and soil properties in brackish tidal marsh of the Mullica River, New Jersey. *Estuaries*, 22, 927–935.

Yuhas, C.E. (2001). *Benthic communities in Spartina alterniflora and Phragmites australis dominated salt marshes*. Master's thesis, New Jersey Institute of Technology/Rutgers University, New Brunswick, NJ.

## Glossary

**Amphipod:** A small crustacean of the order Amphipoda.

**Analysis of variance (ANOVA):** Statistical method that yields values that can be tested to determine whether a significant relation exists between variables.

**Benthic:** Organisms (e.g., protozoa, nematodes) living in sediments on sea or lake bottoms.

**Decapod:** A ten-legged crustacean of the order Decapoda (e.g., crab, lobster).

**df (degrees of freedom) :** In statistics, degrees of freedom are the number of values in probability distributions that are free to be varied.

**Epibenthic:** Organisms living on the surface of sediments on sea or lake bottoms.

**Infauna:** Benthic organisms (see above) that dig into the sediment bed or construct tubes or burrows.

**Log-base-10 transformed (logarithmic transformation):** Statistical technique whereby each observation or raw value of data is replaced by the log of itself.

**Meiofauna:** Organisms that are larger than microfauna and smaller than macrofauna. They will generally pass through a 1-mm mesh but not a 0.3-mm mesh.

**Mesohaline:** Of or relating to a body of water with a salinity measure between 5 and 18 parts per thousand.

**Mitigation (Mitigation banking):** The process of preserving, enhancing, restoring, or creating habitat to compensate for (current or future) habitat disturbances elsewhere, especially due to development.

**Multiple analysis of variance (MANOVA):** An extension of ANOVA (see above) to cover cases where there is more than one dependent variable.

**Nekton:** Actively swimming marine or freshwater organisms.

**Oligochaetes:** Any of various annelid worms of the class Oligochaeta, including the earthworms and a few small freshwater forms.

**Oligohaline:** Of or relating to a body of water with a salinity measure of less than 5 parts per thousand (or 5 grams of salt per liter).

**p < 0.05:** An indicator of statistical significance in which the probability of the result of a study being a chance occurrence is less than 5 in 100.

**Polychaetes:** Any of various annelid worms of the class Polychaeta, including mostly marine worms such as the lugworm, characterized by fleshy-paired appendages tipped with bristles on each body segment.

**Refractometer:** An instrument that measures the composition of liquids using light refraction.

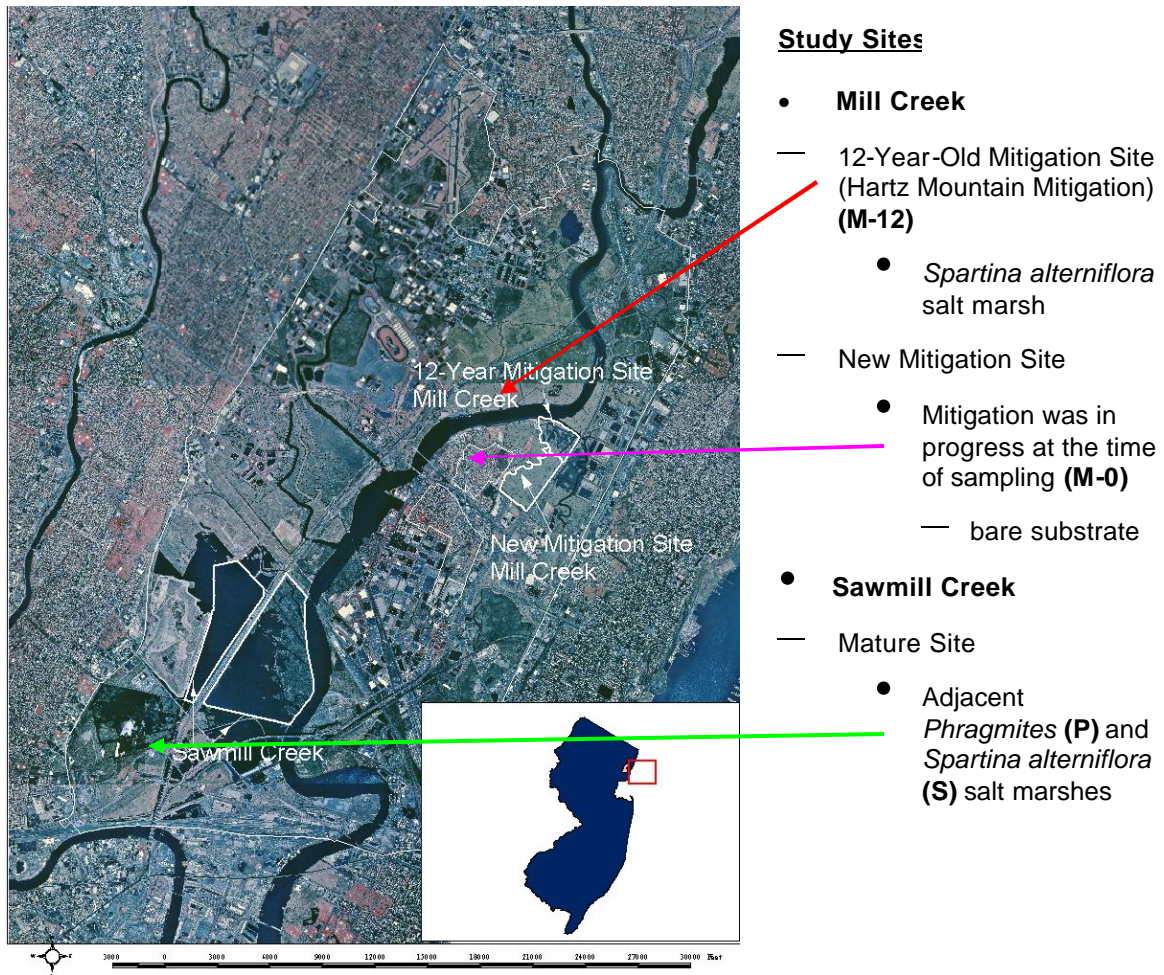
**Richness:** The number of species in an area.

**Stable isotope:** Any naturally occurring, nondecaying isotope (see above) of an element. Many elements have several stable isotopes. For example, carbon (C) has carbon 12 (<sup>12</sup>C) and carbon 13 (<sup>13</sup>C).

**Student-Newman-Keuls (SNK) multiple comparison test:** A statistical method for determining differences among groups of samples.

**Taxon (plural: taxa):** A taxonomic rank, such as family, genus, or species.

Figure 1. Site map of Hackensack Meadowlands District, New Jersey



**Figure 2. Sawmill Creek—adjacent stands of *Phragmites australis* (P) and *Spartina alterniflora* (S).**





**Figure 3. Mill Creek—12-year-old mitigation site (M-12).**



**Figure 4.: Mill Creek—new mitigation site (M-0).**



Figure 5. Mean percentage of silt (horizontal lines), sand (dotted pattern), and clay (vertical lines) at the creek bank and edge of the vegetation habitats of the Sawmill Creek natural sites. Samples were taken at Sawmill Creek—*Spartina alterniflora* (S) and Sawmill Creek—*Phragmites australis* (P) during September 1999.

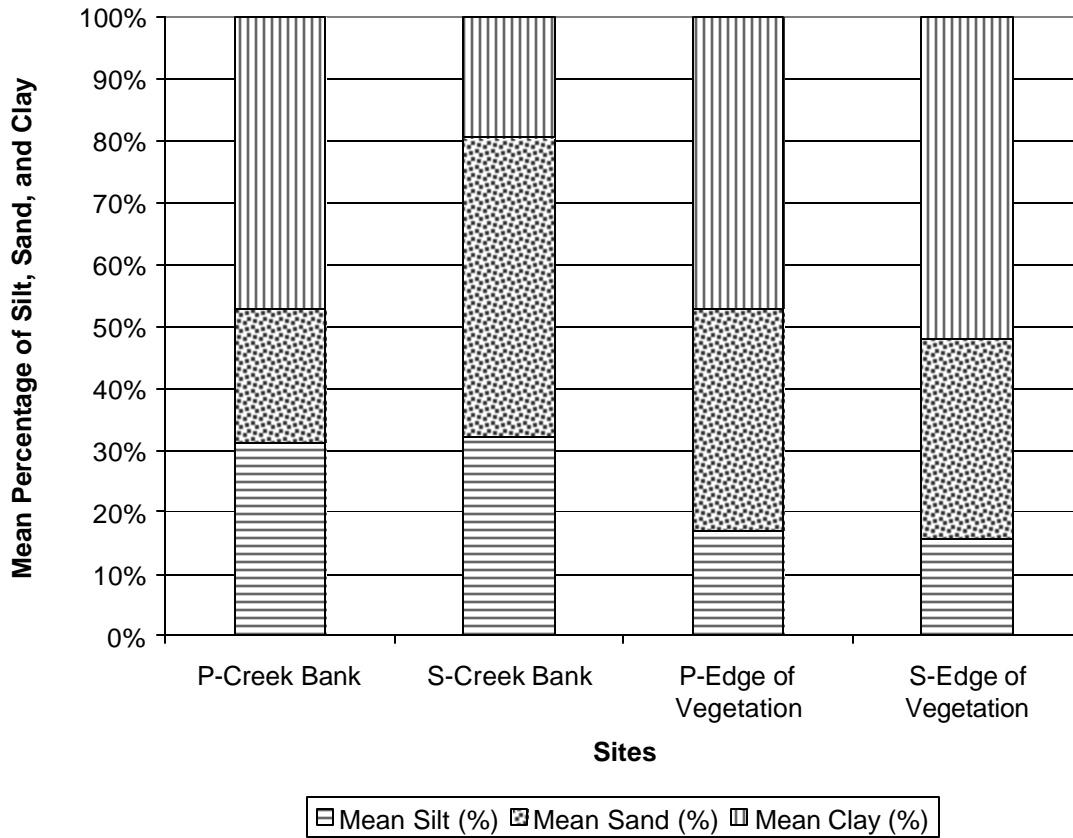


Figure 6. Mean abundance (#/m<sup>2</sup>) of benthic invertebrates at the creek-bank habitat and the edge of the vegetation habitats. Sampled at Sawmill Creek—*S. alterniflora* (S) and Sawmill Creek—*P. australis* (P) from June to September 1999.

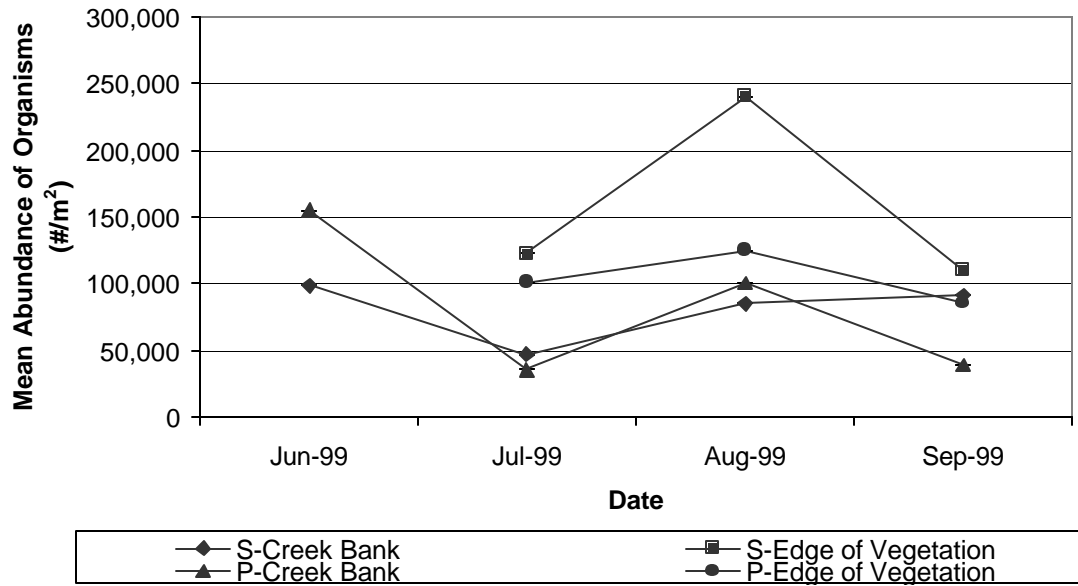
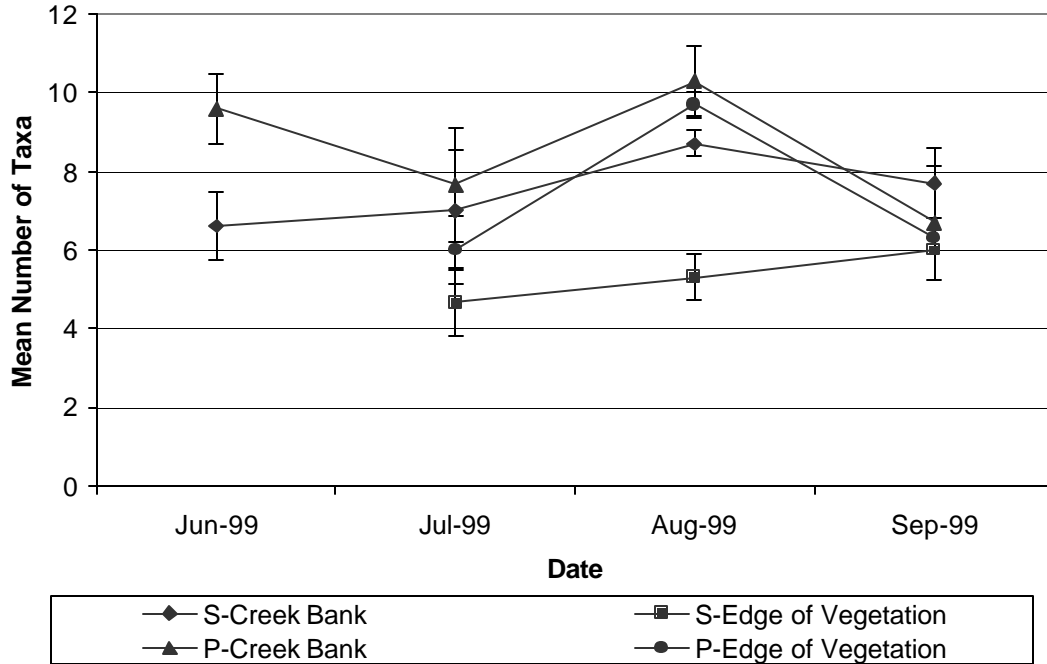
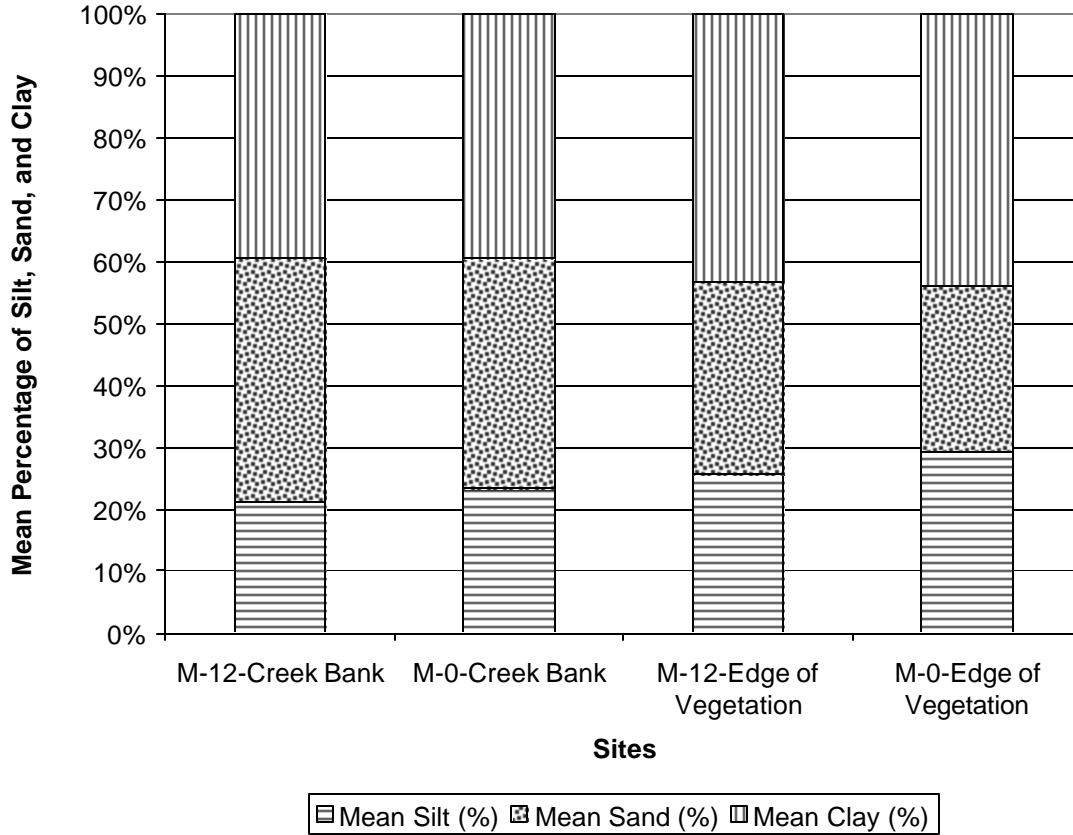


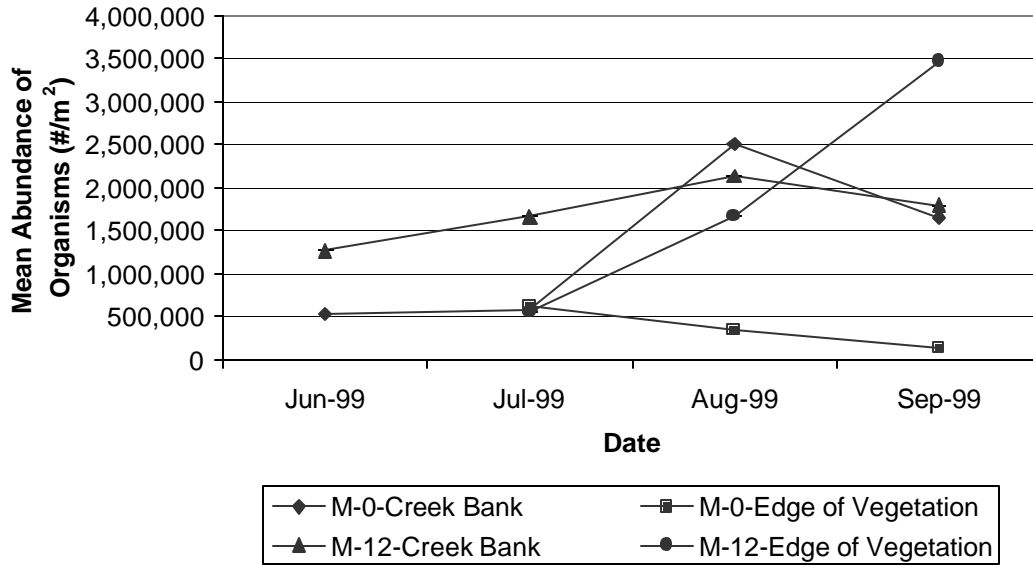
Figure 7. Mean taxa richness at the creek bank and the edge of vegetation habitats. Samples were taken at Sawmill Creek—*S. alterniflora* (S) and Sawmill Creek—*P. australis* (P) from June to September 1999.



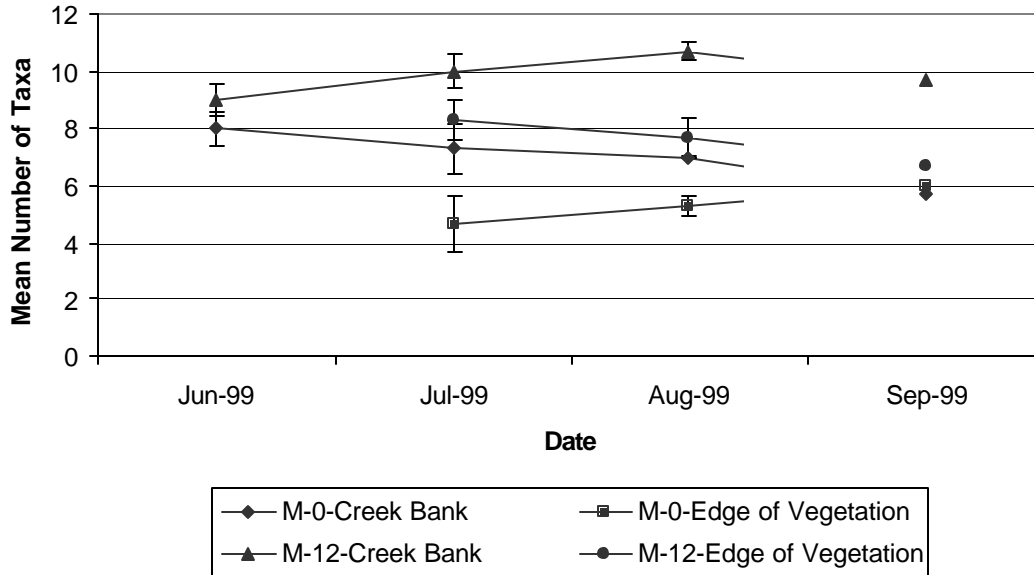
**Figure 8. Mean percentage of silt, sand, and clay at the Mill Creek mitigation sites at the creek bank and edge of the vegetation habitats. Samples were taken at the 12-year-old mitigation site (M-12) and new mitigation site (M-0) during September 1999.**



**Figure 9. Mean abundance (#/m<sup>2</sup>) of benthic invertebrates between the creek bank and the edge of the vegetation habitats. Samples were taken at the 12-year-old mitigation site (M-12) and new mitigation site (M-0) from June to September 1999.**



**Figure 10. Mean taxa richness at the creek bank and edge of vegetation habitats. Samples were taken at the 12-year-old mitigation site (M-12) and new mitigation site (M-0) from June to September 1999.**





**Table 1. Mean percentage of organic matter, silt, sand, and clay at Sawmill Creek—*P. australis* (P), Sawmill Creek—*S. alterniflora* (S), 12-year-old mitigation site—Mill Creek (M-12), and new mitigation site—Mill Creek (M-0).**

Site	Mean Percentage of Organic Matter +/- Standard Error	Mean Percentage of Silt +/- Standard Error	Mean Percentage of Sand +/- Standard Error	Mean Percentage of Clay +/- Standard Error
P-Creek Bank	1.582+/-0.07	31.1+/-1.1	21.8+/-3.63	47.1+/-2.57
P-Edge of Vegetation	2.281+/-0.04	17.1+/-0.2	35.8+/-2.47	47.1+/-2.56
S-Creek Bank	3.084+/-0.25	32.2+/-4.0	48.2+/-7.14	19.5+/-6.89
S-Edge of Vegetation	2.157+/-0.08	15.8+/-2.7	32.2+/-5.53	52.0+/-3.16
M-12-Creek Bank	1.332+/-0.02	21.1+/-1.1	39.6+/-4.25	39.3+/-3.31
M-12-Edge of Vegetation	2.016+/-0.14	25.6+/-11.0	31.1+/-9.10	43.3+/-5.07
M-0-Creek Bank	1.688+/-0.09	23.3+/-1.2	37.1+/-2.32	39.5+/-3.47
M-0-Edge of Vegetation	2.312+/-0.08	29.3+/-0.7	26.7+/-3.84	44.0+/-4.43

**Table 2. P values and degrees of freedom for organic matter, silt, sand, and clay at Sawmill Creek—*P. australis* (P), Sawmill Creek—*S. alterniflora* (S), 12-year-old mitigation site—Mill Creek (M-12), and new mitigation site—Mill Creek (M-0).**

Site	Organic Matter	Silt	Sand	Clay
P-Creek Bank				
P-Edge of Vegetation	P=0.0656/df=1	P=0.0002/df=1	P=0.0047/df=1	P=1.0000/df=1
S-Creek Bank				
S-Edge of Vegetation	P=0.0251/df=1	P=0.0273/df=1	P=0.6548/df=1	P=0.0128/df=1
P-Creek Bank				
S-Creek Bank	P=0.0003/df=2	P=0.5880/df=2	P=0.0038/df=2	P=0.0051/df=2
P-Edge of Vegetation				
S-Edge of Vegetation	P=0.7685/df=1	P=0.6587/df=1	P=0.2943/df=1	P=0.2943/df=1
M-12-Creek Bank				
M-12-Edge of Vegetation	P=0.0088/df=1	P=0.7057/df=1	P=0.5997/df=1	P=0.5451/df=1
M-0-Creek Bank				
M-0-Edge of Vegetation	P=0.0075/df=1	P=0.0106/df=1	P=0.2397/df=1	P=0.4719/df=1
M-12-Creek Bank				
M-0-Creek Bank	P=0.0027/df=2	P=0.0177/df=2	P=0.0042/df=2	P=0.0026/df=2
M-12-Edge of Vegetation				
M-0-Edge of Vegetation	P=0.1468/df=1	P=0.7487/df=1	P=0.9259/df=1	P=0.9259/df=1

**Table 3. Mean abundance (#/m<sup>2</sup>), mean taxa richness, and mean Shannon-Wiener Diversity Index at Sawmill Creek—*P. australis* (P), Sawmill Creek—*S. alterniflora* (S), 12-year-old mitigation site—Mill Creek (M-12), and new mitigation site—Mill Creek (M-0).**

Site	Mean Abundance +/- Standard Error (#/m <sup>2</sup> )	Mean Taxa Richness +/- Standard Error	Mean Shannon-Wiener Diversity Index +/- Standard Error
<b>June</b>			
P-Creek Bank	155030+/-15.52	9.67+/-0.88	1.66+/-0.06
S-Creek Bank	98297+/-26.87	6.67+/-0.88	1.33+/-0.08
M-12-Creek Bank	1262279+/-511.38	9.00+/-0.58	1.05+/-0.18
M-0-Creek Bank	528778+/-57.36	8.00+/-0.58	1.12+/-0.13
<b>July</b>			
P-Creek Bank	35447+/-15.34	7.68+/-1.45	1.64+/-0.14
P-Edge of Vegetation	100811+/-30.61	6.00+/-0.58	1.06+/-0.22
S-Creek Bank	46928+/-28.83	7+/-1.53	1.50+/-0.18
S-Edge of Vegetation	122097+/-31.54	4.67+/-0.67	0.71+/-0.12
M-12-Creek Bank	1660916+/-622.00	10.00+/-0.58	0.94+/-0.17
M-12-Edge of Vegetation	554756+/-49.11	8.33+/-0.33	1.13+/-0.02
M-0-Creek Bank	578220+/-106.29	7.33+/-0.88	0.79+/-0.09
M-0-Edge of Vegetation	611991+/-158.60	5.67+/-0.33	0.94+/-0.09
<b>August</b>			
P-Creek Bank	100309+/-30.59	10.33+/-0.88	1.77+/-0.03
P-Edge of Vegetation	125113+/-36.43	9.67+/-0.88	1.37+/-0.08
S-Creek Bank	84889+/-41.38	8.67+/-0.33	1.65+/-0.14
S-Edge of Vegetation	240506+/-80.48	5.33+/-0.88	1.06+/-0.07
M-12-Creek Bank	2139414+/-720.27	10.67+/-0.33	0.66+/-0.08
M-12-Edge of Vegetation	1670721+/-608.09	7.67+/-0.67	0.80+/-0.10
M-0-Creek Bank	2508134+/-574.03	7.00+/-0.00	0.37+/-0.06
M-0-Edge of Vegetation	343329+/-71.01	7.00+/-1.00	1.33+/-0.10
<b>September</b>			

P-Creek Bank	38799+/-20.38	6.67+/-1.45	1.49+/-0.19
P-Edge of Vegetation	85225+/-6.57	6.33+/-0.33	1.33+/-0.14
S-Creek Bank	91342+/-14.47	7.67+/-0.88	1.30+/-0.06
S-Edge of Vegetation	109778+/-3.46	6.00+/-0.58	1.11+/-0.14
M-12-Creek Bank	1791393+/-446.71	9.67+/-0.67	0.63+/-0.13
M-12-Edge of Vegetation	3470745+/-506.24	6.67+/-0.67	0.33+/-0.08
M-0-Creek Bank	1652536+/-119.98	5.67+/-0.33	0.38+/-0.02
M-0-Edge of Vegetation	128801+/-4.91	6.67+/-0.33	1.31+/-0.10

**Table 4. P values and degrees of freedom for abundance, taxa richness, and Shannon-Wiener Diversity Index at Sawmill Creek—*P. australis* (P), Sawmill Creek—*S. alterniflora* (S), 12-year-old mitigation site—Mill Creek (M-12), and new mitigation site—Mill Creek (M-0).**

Site	Abundance	Taxa Richness	Shannon-Wiener Diversity Index
<b>June</b>			
P-Creek Bank			
S-Creek Bank	P=0.0962/df=1	P=0.0739/df=1	P=0.0347/df=1
M-12-Creek Bank			
M-0-Creek Bank	P=0.1487/df=1	P=0.2879/df=1	P=0.7384/df=1
<b>July</b>			
P-Creek Bank			
P-Edge of Vegetation	P=0.0845/df=1	P=0.3465/df=1	P=0.1002/df=1
S-Creek Bank			
S-Edge of Vegetation	P=0.1061/df=1	P=0.2341/df=1	P=0.0218/df=1
P-Creek Bank			
S-Creek Bank	P=0.8046/df=1	P=0.7676/df=1	P=0.5818/df=1
P-Edge of Vegetation			
S-Edge of Vegetation	P=0.5908/df=1	P=0.2051/df=1	P=0.2410/df=1
M-12-Creek Bank			
M-12-Edge of Vegetation	P=0.0357/df=1	P=0.0668/df=1	P=0.1653/df=1
M-0-Creek Bank			
M-0-Edge of Vegetation	P=0.8881/df=1	P=0.1518/df=1	P=0.2609/df=1
M-12-Creek Bank			
M-0-Creek Bank	P=0.0502/df=1	P=0.0647/df=1	0.4636/df=1
M-12-Edge of Vegetation			
M-0-Edge of Vegetation	P=0.8001/df=1	P=0.0048/df=1	P=0.0333/df=1
<b>August</b>			
P-Creek Bank			
P-Edge of Vegetation	P=0.5634/df=1	P=0.6213/df=1	P=0.0081/df=1
S-Creek Bank			
S-Edge of Vegetation	P=0.0962/df=1	P=0.0241/df=1	P=0.0189/df=1
P-Creek Bank			
S-Creek Bank	P=0.6425/df=1	P=0.1518/df=1	P=0.4453/df=1
P-Edge of Vegetation	P=0.1959/df=1	P=0.0255/df=1	P=0.0371/df=1

S-Edge of Vegetation			
M-12-Creek Bank			
M-12-Edge of Vegetation	P=0.6105/df=1	P=0.0158/df=1	P=0.3445/df=1
M-0-Creek Bank			
M-0-Edge of Vegetation	P=0.0024/df=1	P=1.0000/df=1	P=0.0012/df=1
M-12-Creek Bank			
M-0-Creek Bank	P=0.6110/df=1	P=0.0004/df=1	P=0.0464/df=1
M-12-Edge of Vegetation			
M-0-Edge of Vegetation	P=0.0133/df=1	P=0.6087/df=1	P=0.0225/df=1
<b>September</b>			
P-Creek Bank			
P-Edge of Vegetation	P=0.1226/df=1	P=0.8340/df=1	P=0.5500/df=1
S-Creek Bank			
S-Edge of Vegetation	P=0.2151/df=1	P=0.1890/df=1	P=0.2565/df=1
P-Creek Bank			
S-Creek Bank	P=0.1149/df=1	P=0.5879/df=1	P=0.4242/df=1
P-Edge of Vegetation			
S-Edge of Vegetation	P=0.0212/df=1	P=0.6433/df=1	P=0.3270/df=1
M-12-Creek Bank			
M-12-Edge of Vegetation	P=0.0601/df=1	P=0.0335/df=1	P=0.1173/df=1
M-0-Creek Bank			
M-0-Edge of Vegetation	P<0.0001/df=1	P=0.1012/df=1	P=0.0007/df=1
M-12-Creek Bank			
M-0-Creek Bank	P=0.8974/df=1	P=0.0058/df=1	P=0.1371/df=1
M-12-Edge of Vegetation			
M-0-Edge of Vegetation	P<0.0001/df=1	P=1.0000/df=1	P=0.0014/df=1

**Table 5. Taxa present at (+) or absent from (–) Sawmill Creek—*P. australis* (P), Sawmill Creek—*S. alterniflora* (S), 12-year-old mitigation site—Mill Creek (M-12), and new mitigation site—Mill Creek (M-0); sampled from June to September 1999.**

Site	Abundance	Taxa Richness	Shannon-Wiener Diversity Index
<b>June</b>			
P-Creek Bank S-Creek Bank	P=0.0962/df=1	P=0.0739/df=1	P=0.0347/df=1
M-12-Creek Bank M-0-Creek Bank	P=0.1487/df=1	P=0.2879/df=1	P=0.7384/df=1
<b>July</b>			
P-Creek Bank P-Edge of Vegetation	P=0.0845/df=1	P=0.3465/df=1	P=0.1002/df=1
S-Creek Bank S-Edge of Vegetation	P=0.1061/df=1	P=0.2341/df=1	P=0.0218/df=1
P-Creek Bank S-Creek Bank	P=0.8046/df=1	P=0.7676/df=1	P=0.5818/df=1
P-Edge of Vegetation S-Edge of Vegetation	P=0.5908/df=1	P=0.2051/df=1	P=0.2410/df=1
M-12-Creek Bank M-12-Edge of Vegetation	P=0.0357/df=1	P=0.0668/df=1	P=0.1653/df=1
M-0-Creek Bank M-0-Edge of Vegetation	P=0.8881/df=1	P=0.1518/df=1	P=0.2609/df=1
M-12-Creek Bank M-0-Creek Bank	P=0.0502/df=1	P=0.0647/df=1	0.4636/df=1
M-12-Edge of Vegetation M-0-Edge of Vegetation	P=0.8001/df=1	P=0.0048/df=1	P=0.0333/df=1
<b>August</b>			
P-Creek Bank	P=0.5634/df=1	P=0.6213/df=1	P=0.0081/df=1

P-Edge of Vegetation			
S-Creek Bank			
S-Edge of Vegetation	P=0.0962/df=1	P=0.0241/df=1	P=0.0189/df=1
P-Creek Bank			
S-Creek Bank	P=0.6425/df=1	P=0.1518/df=1	P=0.4453/df=1
P-Edge of Vegetation			
S-Edge of Vegetation	P=0.1959/df=1	P=0.0255/df=1	P=0.0371/df=1
M-12-Creek Bank			
M-12-Edge of Vegetation	P=0.6105/df=1	P=0.0158/df=1	P=0.3445/df=1
M-0-Creek Bank			
M-0-Edge of Vegetation	P=0.0024/df=1	P=1.0000/df=1	P=0.0012/df=1
M-12-Creek Bank			
M-0-Creek Bank	P=0.6110/df=1	P=0.0004/df=1	P=0.0464/df=1
M-12-Edge of Vegetation			
M-0-Edge of Vegetation	P=0.0133/df=1	P=0.6087/df=1	P=0.0225/df=1
<b>September</b>			
P-Creek Bank			
P-Edge of Vegetation	P=0.1226/df=1	P=0.8340/df=1	P=0.5500/df=1
S-Creek Bank			
S-Edge of Vegetation	P=0.2151/df=1	P=0.1890/df=1	P=0.2565/df=1
P-Creek Bank			
S-Creek Bank	P=0.1149/df=1	P=0.5879/df=1	P=0.4242/df=1
P-Edge of Vegetation			
S-Edge of Vegetation	P=0.0212/df=1	P=0.6433/df=1	P=0.3270/df=1
M-12-Creek Bank			
M-12-Edge of Vegetation	P=0.0601/df=1	P=0.0335/df=1	P=0.1173/df=1
M-0-Creek Bank			
M-0-Edge of Vegetation	P<0.0001/df=1	P=0.1012/df=1	P=0.0007/df=1



M-12-Creek Bank			
M-0-Creek Bank	P=0.8974/df=1	P=0.0058/df=1	P=0.1371/df=1
M-12-Edge of Vegetation			
M-0-Edge of Vegetation	P<0.0001/df=1	P=1.0000/df=1	P=0.0014/df=1

Note: Sawmill Creek and Mill Creek sites should not be compared, even though they are both represented in this table.

**Table 6. Totals and percent totals of the common and uncommon species.**

<b>Sawmill Creek—<i>Phragmites australis</i></b>										
<b>Common Species</b>	<i>Cyanthura polita</i>	Ceratopogonidae	Nereidae	Spionidae	<i>Hobsonia florida</i>	<i>Streblospio benedicti</i>	Copepoda	<i>Manayunkia aestuarina</i>	Nematoda	Oligochaeta
Total	23	26	57	75	121	138	296	386	526	559
% of Total	1.00	1.13	2.48	3.27	5.27	6.02	12.90	16.83	22.93	24.37
<b>Uncommon Species</b>	<i>Macoma bathica</i>	<i>Anurida martina</i>	Acarina	<i>Edotea triloba</i>	Chironomidae	Foraminifera	Turbellaria	Insect Larvae		
Total	3	4	5	5	6	14	19	22		
% of Total	0.13	0.17	0.22	0.22	0.26	0.61	0.83	0.96		
<b>Sawmill Creek—<i>Spartina alterniflora</i></b>										
<b>Common Species</b>	Spionidae	<i>Hobsonia florida</i>	Nereidae	Copepoda	<i>Streblospio benedicti</i>	Oligochaeta	Nematoda	<i>Manayunkia aestuarina</i>		
Total	30	41	62	126	186	542	873	940		
% of Total	1.05	1.44	2.18	4.43	6.54	19.04	30.67	33.03		
<b>Uncommon Species</b>	Gammarid	Foraminifera	Ceratopogonidae	<i>Cyanthura polita</i>	Turbellaria					
Total	3	7	9	10	11					
% of Total	0.11	0.25	0.32	0.35	0.39					
<b>New Mitigation Site—Mill Creek</b>										
<b>Common Species</b>	Gastropoda	Ostracoda	Oligochaeta	Nematoda						
Total	459	1540	2859	17571						
%of Total	2.02	6.77	12.57	77.27						
<b>Uncommon Species</b>	<i>Hobsonia florida</i>	Chironomidae	Copepoda							
Total	31	104	147							
%of Total	0.14	0.46	0.65							
<b>12-yr-old Mitigation Site—Mill Creek</b>										
<b>Common Species</b>	<i>Manayunkia aestuarina</i>	Copepoda	Oligochaeta	Nematoda						
Total	826	1260	6168	35853						
%of Total	1.84	2.80	13.73	79.78						
<b>Uncommon Species</b>	Gastropoda	Ostracoda								
Total	273	362								
%of Total	0.61	0.81								